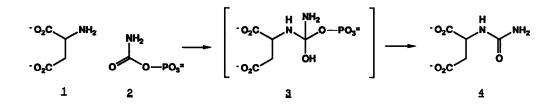
SYNTHESIS OF POTENTIAL INHIBITORS OF THE ENZYME ASPARTATE TRANSCARBAMOYLASE 1

Stephen D. Lindell * and Richard M. Turner

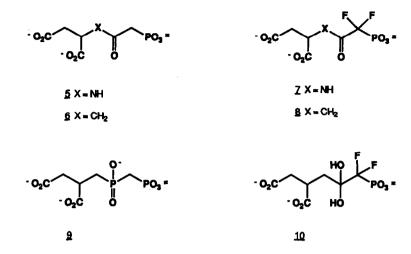
Schering Agrochemicals Limited, Chesterford Park Research Station, Saffron Walden, Essex, CB10 1XL, ENGLAND

<u>ABSTRACT</u>: Two difluoromethylenephosphonates, 7 and 8, and the phosphinylmethylenephosphonate 9 were prepared as potential inhibitors of aspartate transcarbamoylase (ATCase). The synthesis of compound 8 was facilitated by development of a new direct method for converting nitroalkanes to alkanoic acids. Of the three phosphonates, the amide 7 proved to be a good inhibitor of mung bean ATCase $(I_{50} ~ 5\mu M)$.

Aspartate transcarbamoylase (EC 2.1.3.2., ATCase) catalyses the condensation of L-aspartate $(\underline{1})$ with carbamoyl phosphate $(\underline{2})$ to give N-carbamoyl-L-aspartate $(\underline{4})$. This is the first committed step in <u>de novo</u> pyrimidine biosynthesis, and inhibitors of this enzyme are of potential interest both as agrochemicals² and pharmaceuticals.³



N-(Phosphonoacetyl)-L-aspartate, PALA $(5)^4$ and (4,5-dicarboxy-2-ketopentyl)phosphonate, DIKEP $(6)^5$ are both potent inhibitors of ATCase $(K_i$ values about 10^{-8} M and 10^{-7} M, respectively). These compounds are multi-substrate analogues,⁶ which may also exhibit some transition state characteristics.³ In both PALA (5) and DIKEP (6) a methylenephosphonate group has been used as a stable replacement for a phosphate moiety. Although this is a good isosteric replacement, the difference in electronegativity between oxygen and methylene carbon means it is less good electronically. Work by Blackburn et al⁷ led us to postulate that this electronic disparity would be overcome in the diffuoromethylenephosphonate analogues 7 and 8, thereby leading to tighter enzyme binding. In addition, the highly electrophilic nature of the diffuoroketone 8 means that it should readily react with a water molecule (possibly one of those present in the active site⁸) to give the hydrate 10. Both 10 and the phosphinate 9 can be regarded as mimics of the high energy tetrahedral reaction intermediate 3. "Transition state" inhibitors based on α -fluoroketones or tetrahedral phosphorus are well documented, and have the potential for very tight binding.⁹ This communication describes the synthesis and ATCase-inhibitory properties of the three phosphonates 7, 8 and 9.



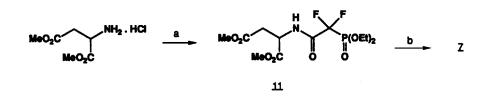
The amide $\underline{11}^{10}$ ($[\alpha]_{p}^{25}$ 39°(c 2, CHCl₃)) was synthesised by coupling L-aspartic acid dimethyl ester hydrochloride with diethyl difluorophosphonoacetic acid¹¹ (Scheme I). De-esterification with trimethylsilyl bromide¹² followed by hydroxide yielded the required amide <u>7</u>, isolated as the tetra-lithium salt monohydrate ($[\alpha]_{p}^{25}$ 3.6° (c 2, H₂0)).¹⁰

The ketone $\underline{8}$ was synthesised as outlined in Scheme II. Michael addition of nitromethane to diethyl itaconate yielded the nitro compound $\underline{12}$.¹⁰ This material was cleanly converted by basic buffered KMnO₄ into the carboxylic acid $\underline{13}$.^{10,13} This reaction is noteworthy in that although KMnO₄ has been used to convert nitroalkanes to aldehydes,¹⁴ and aldehydes to acids,¹⁵ it has not been previously reported as a reagent for direct conversion. In fact we have been able to find very few alternative mild methods for achieving this transformation.¹⁶ Our method tolerates a variety of functional groups (ester, amide, primary alcohol, benzylic methylene) and generally gives analytically pure products without chromatography. The acid 13 was converted with oxalyl chloride¹⁷ into the acid chloride 14, which was directly coupled with [(diethylphosphinyl)difluoromethyl]zinc bromide¹⁸ to yield <u>15</u>.¹⁰ De-esterification as before yielded the ketone <u>8</u> (again isolated as the tetralithium salt monohydrate)¹⁰ in 42**X** overall yield.

The synthesis of the final target, the phosphinate 9, was achieved as shown in Scheme III. Michael addition of ethyl hypophosphite¹⁹ to diethyl itaconate yielded the phosphinate $\frac{16}{17}$.¹⁰ The sodium salt of $\frac{16}{16}$ was condensed with diethyl phosphonomethyltriflate^{20,21} to yield $\frac{17}{17}$.¹⁰ This reaction is of interest in that, coupled with recent advances in the synthesis of P-H phosphinates,²² it provides a new method for preparing the biologically interesting phosphinylmethylenephosphonates.²³ Finally, acid catalysed hydrolysis yielded the phosphinate 9, isolated as the pentasodium salt dihydrate.¹⁰

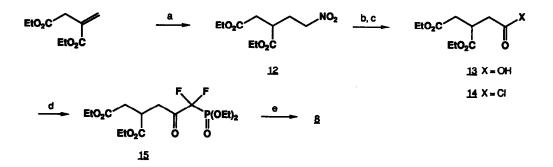
Compounds 7, 8 and 9 were tested for activity against mung bean ATCase.²⁴ Unfortunately, both the difluoroketone 8 and the phosphinate 9 were inactive at the highest concentrations measured (100 μ M). This may be because ATCase catalyses via a "closed transition state",²⁵ so that it can only bind inhibitors with a ground state trigonal carbonyl. The difluoroketone 8 probably exists as the hydrate 10 under the assay conditions,²⁶ and is thus unable to bind. Gratifyingly, the amide 7 was a good inhibitor of ATCase (I₅₀ - 5 μ M), though it was less tightly bound than PALA 5 (I₅₀ - 1 μ M). It has been suggested that the carbonyl group of PALA is protonated at the active site of ATCase.²⁷ This may, at least in part, explain the difference in inhibition levels of 5 and 7. None of the compounds exhibited any significant agrochemical activity.

<u>ACKNOWLEDGEMENTS</u>: We wish to thank Drs A.C. Baillie, C.G. Earnshaw, K. Wright and the late Mr B.J. Wright of Schering for helpful discussions during the course of this work. We also wish to thank Mr. J. Cant for performing the enzyme assays.



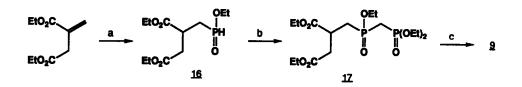
(a) 1eq (EtO) P(O)CF₂CO₂H, 1eq 1-hydroxybenzotriazole, 1.1eq DCC, CH₂Cl₂, r.t., 2 days; 46%.
(b) (i) 5eq TMSBr, r.t., 5h; (ii) 1M NaOH, r.t., 5h; (iii) 'Amberlite' IR-120, H⁺ form; (iv) 4eq LiOH; 98%.

SCHEME II



(a) 40eq CH₂NO₂, 1eq NaOEt, EtOH, Δ , 5h; 73%. (b), to give <u>13</u>: (i) 2eq KOH, 5eq K₂HPO₄, <u>tBuOH</u>, H₂O then 4eq KMnO₄, 25°C, 30 min; (ii) sat'd Na₂SO₃; (iii) 2M HCl; 96%. (c) to give <u>14</u>: 1.05 eq (COCl)₂, PhMe, 40°C, 3 h; quant. (d) 1.05 eq (EtO)₂P(O)CF₂ZnBr, 0.08 eq CuBr, DME, 0°C+rt, 20h; 60%. (e) (i) 5 eq TMSBr, r.t., 3 days; (ii) 1M NaOH, r.t., 5h; (iii) 'Amberlite' IR-120, H⁺ form; (iv) 4eq LiOH; 98%.

SCHEME III



(a) 1eq H₂P(0)OEt, EtOH, 80°C, 24 h; 41%. (b) 1eq (EtO)₂P(0)CH₂OSO₂CF₃, 1eq NaH, DME, r.t., 80 min; 74%. (c) (i) 6M HCl ,Δ, 16 h; (ii) 5eq NaOH; 73%.

REFERENCES AND NOTES

1	This work is part of an ongoing interest at Schering in using biochemical reasoning to generate leads for the discovery of new agrochemicals. See A.C. Baillie, K. Wright, B.J. Wright, and C.G. Earnshaw, <u>Pest. Biochem. Physiol</u> ., 1988, <u>30</u> , 103 and references therein.
2	a). A.C. Baillie, B.J. Wright and K. Wright, Schering internal report (1977). b). P. Böger, J. Pesticide Sci., 1987, 12, 749.
3	G. R. Stark and P. A. Bartlett, Pharmac. Ther., 1983, 23, 45.
4	K. D. Collins and G. R. Stark, <u>J. Biol. Chem.</u> , 1971, <u>246</u> , 6599.
5	E. A. Swyryd, S. S. Seaver and G. R. Stark, J. Biol. Chem., 1974, 249, 6945.
6	L. Frick and R. Wolfenden in, "Design of Enzyme Inhibitors as Drugs", M. Sandler and
•	H. J. Smith, eds., Oxford Univ. Press, Oxford, 1989, 19.
7	G. M. Blackburn, D. A. England and F. Kolkmann, J. Chem. Soc. Chem. Comm., 1981, 930.
8	K. L. Krause, K. W. Volz and W. N. Lipscomb, J. Mol. Biol., 1987, 193, 527.
9	J. V. Schloss in "Target Sites of Herbicide Action", P. Böger and G. Sandmann, eds.,
2	CRC Press Inc., Boca Raton, 1989, 165.
10	¹ H NMR and IR spectra are consistent with the assigned structure. A satisfactory
10	compound.
11	G. M. Blackburn, D. Brown and S. J. Martin, <u>J. Chem. Res. (S)</u> , 1985, 92.
12	C. E. McKenna and J. Schmidhauser, J. Chem. Soc. Chem. Comm., 1979, 739.
13	To a stirred solution of nitroalkane 12 (5.0 g, 20.2 mmol) in tBuOH (100 ml) at
	25°C was added an aqueous solution 0.5 M in KOH and 1.25 M in K, HPO, (80 ml). The
	yellow solution was stirred for ca 0.5 min, and than an aqueous 0.5 M solution of
	$KMnO_4$ (160 ml) was added. After 30 min the reaction was quenched by addition of
	saturated aqueous Na ₂ SO ₃ (100 ml), and the pH was adjusted to <u>ca</u> 3 with 2M HCl to
	give a clear reaction solution. Extraction with EtOAc (5 x 150 ml), drying (MgSO ₄)
	and evaporation in vacuo yielded acid 13 as a microanalytically pure pale yellow
	oil (4.52 g, 19.5 mmol, 96%).
14	N. Kornblum, A. S. Erickson, W. J. Kelly and B. J. Henggleler, J. Org. Chem., 1982,
	47, 4534. K. Steliou and MA. Poupart, ibid., 1985, 50, 4971.
15	A. Abiko, J. C. Roberts, T. Takemasa and S. Masamune, Tetrahedron Letters, 1986,
	27, 4537.
16	G. Calderari and D. Seebach, <u>Helv. Chim. Acta</u> , 1985, <u>68</u> , 1592.
17	J. E. H. Hancock and R. P. Linstead, <u>J. Chem. Soc</u> ., 1953, 3490.
18	D. J. Burton and L. G. Sprague, J. Org. Chem., 1988, 53, 1523. The zinc metal was
	activated as described by P. Knochel, M. C. P. Yeh, S. C. Berk and J. Talbert,
	<u>ibid</u> , 2390.
19	S. J. Fitch, <u>J. Am. Chem. Soc</u> ., 1964, <u>86</u> , 61. M. J. Gallagher and J. Sussman,
	Phosphorus, 1975, 5, 91.
20	D. P. Phillion and S. S. Andrew, Tetrahedron Letters, 1986, 27, 1477.
21	NaH (80% oil dispersion, 0.15 g, 5.0 mmol) was added to a stirred solution of the
	phosphinate 16 (1.4 g, 5.0 mmol) and diethyl phosphonomethyltriflate (1.5 g, 5.0
	mmol) in dry DME (5 ml) under nitrogen at r.t. After 80 min, the mixture was
	poured into sat'd aq. NH ₄ Cl (50 ml), extracted with CH ₂ Cl ₂ (2 x 50 ml), dried
	(MgSO ₄) and evaporated. Flash chromatography yielded the phosphinate 17 as a
~~	colourless oil (1.60 g, 3.7 mmol, 74%).
22	R. Engel and S. Chakraborty, Synth. Comm., 1988, 18, 665. J. G. Dingwall, J.
	Ehrenfreund and R. G. Hall, Tetrahedron, 1989, 45, 3787. E. A. Boyd, M. Corless,
12	K. James and A. C. Regan, <u>Tetrahedron Letters</u> , 1990, <u>31</u> , 2933.
23	M. H. B. Stowell, J. F. Witte and R. W. McClard, <u>Tetrahedron Letters</u> , 1989, <u>30</u> , 411 and references therein.
24	B. S. Achar, H. S. Savithri, C. S. Vaidyanathan and N. Appaji Rao, <u>Eur. J.</u>
24	Biochem., 1974, 47 , 15. Enzyme from a 45 - 60% (NH ₄) ₂ SO ₄ precipitate was
	preincubated with 16 mM aspartate, inhibitor was added and the reaction was started
	with 1 mM carbamoyl phosphate.
25	R. Kluger and T. Smyth, J. Am. Chem. Soc., 1981, 103, 1214.
26	M.H. Gelb, J.P. Svaren and R.H. Abeles, <u>Biochemistry</u> , 1985, <u>24</u> , 1813.
27	M. F. Roberts, S. J. Opella, M. H. Schaffer, H. M. Phillips and G. R. Stark, <u>J.</u>
<u>.</u>	<u>Biol. Chem</u> ., 1976, <u>251</u> , 5976.